Syntheses of (4,1-Benzoxazepine-3-ylidene)acetic Acid Derivatives and Their Inhibition of Squalene Synthase¹⁾

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The (3,5-trans)-7-chloro-5-(2-chlorophenyl)-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-acetic acid derivatives 1 have been previously identified as potent squalene synthase inhibitors. A series of (4,1-benzoxazepin-3-ylidene)acetic acid derivatives were synthesized and evaluated for their inhibition of rat and human squalene synthase, and the (E)-isomers were found to exhibit potent inhibitory activity, with the same potency as 4,1-benzoxazepine-3-acetic acid derivatives. In contrast the (Z)-isomers did not exhibit significant inhibitory activity, and the active conformation of the 4,1-benzoxazepine-3-acetic acid derivatives was deduced from the folded conformation of the (E)-isomers.

Key words squalene synthase; 4,1-benzoxazepine-3-acetic acid; active conformation; conformationally restricted analogue; (4,1-benzoxazepin-3-ylidene)acetic acid derivative; hypocholestemic agent

Squalene synthase [EC 2.5.1.21] catalyzes the dimerization of farnesyl diphosphate to squalene in cholesterol biosynthesis. This enzymatic step occurs after the pathway branches to other non-steroidal isoprenoids such as dolicol, ubiquinones and isopentenyl t-RNA. Since squalene synthase inhibitors do not interfere with the biosynthesis of these isoprene derivatives, inhibition of this step would arrest only cholesterol biosynthesis and might be useful for the treatment of hyperlipidemia.

In previous papers,¹⁾ we have described chemical modification of (3,5-trans)-7-chloro-5-phenyl-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-acetic acid derivatives and evaluation of their inhibition of squalene synthase. Through the structure-activity relationship study, compounds 1a-c, which have an alkyl group at the 1-position, were found to be potent inhibitors (Fig. 1). In these compounds the conformations of fused ring systems are rigid, but that of the carboxymethyl group at the 3-position, which was essential for potent activity, is very flexible. Thus we considered that restriction of the flexibility of the 3-acetic acid moiety would yield useful information concerning the active conformation of these compounds. We report here the preparation of the (4,1-benzoxazepin-3-ylidene)acetic acid derivatives in which a double bond was introduced between the 3-position and α position (Fig. 1, 2-4) and the evaluation of their inhibition of squalene synthase.²⁾

Fig. 1. Structures of 4,1-Benzoxazepine Derivatives

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Chemistry

The epoxy-amide derivative **6a** was prepared by acylation of the racemic 2-aminobenzylalcohol derivative $5a^{1b}$ with the acid chloride prepared from DL-trans-epoxysuccinic acid monoethyl ester³) and dichloromethylenedimethyliminium chloride.⁴⁾ Intramolucular cyclization⁵⁾ of the epoxy-amide derivative **6a** afforded a *ca.* 1:1 mixture of two products, which were separated by fractional recrystallization to yield the compounds 7a and 7b (Chart 1).⁶⁾ The structures of 7a and 7b were determined by X-ray diffraction analysis, and are shown in Figs. 2 and 3. Both of these compounds are 7membered 4,1-benzoxazepine derivatives and the configurations of 7a and 7b were determined to be $(3RS, 5SR, \alpha SR)$ and $(3RS, 5RS, \alpha SR)$, respectively. The fact that **7a** and **7b** have the same configurations at the 3- and α -positions indicate intramolecular cyclization of 6a proceeded by attack of the oxygen anion on the epoxy carbon bound to the carbamoyl group from the opposite face of the epoxy oxygen (Fig. 4).



Reagents: (a) DL-*trans*-Epoxysuccinic acid monoethyl ester, dichloromethylenedimethyliminium chloride, NaHCO₃, CH₂Cl₂; (b) 1) K₂CO₃, EtOH, 2) fractional recrystallization.

Chart 1

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Fig. 2. Stereoscopic Molecular View of Compound 7a



Fig. 3. Stereoscopic Molecular View of Compound 7b



Fig. 4. Mechanism of Cyclization of 6a

Compound 7a was methanesulfonylated, and the resulting mesylate **8a** was treated with cesium propionate⁷) to yield a mixture of two products (Chart 2). Elimination of the mesyloxy group with sodium acetate instead of cesium propionate proceeded slowly and required excess amount of sodium acetate. Separation of the mixture by column chromatography gave 9a (more polar isomer) and 9b (less polar isomer) in 1:2 ratio. Elemental analysis of these compounds indicated that they have the same composition and mass spectra of both compounds showed a molecular ion peak at m/z 433 (M^+) . Their IR spectra exhibited very similar patterns and in the ¹H-NMR spectra singlet signals due to olefin protons were observed at δ 5.41 and 5.48 ppm of **9a** and **9b**, respectively. On the basis of these results, 9a and 9b were determined to be geometrical isomers of (4,1-benzoxazepin-3-ylidene)acetic acid derivatives. Compounds 9a and 9b were then hydrolyzed to the acids 2a and 2b, respectively and configuration of the double bond was established by X-ray diffraction analysis of compound 2a. As shown in Fig. 5, compound 2a (9a) was determined to be the (E)-isomer, and thus by inference, compound **2b** (**9b**) was the (Z)-isomer.

Compound 7b, which is a diastereoisomer of 7a, could also be converted to 9a, b by mesylation, and subsequent treatment of the resulting 8b with cesium propionate. The ratio of 9a and 9b (1:2) prepared from 7b was identical to that obtained from 7a (Chart 3).⁸⁾



Reagents: (a) MsCl, Et₃N, AcOEt; (b) 1) CH₃CH₂COOCs, DMF, 2) chromatographic separation; (c) 1N NaOH, EtOH.

Chart 2



Fig. 5. Stereoscopic Molecular View of Compound 2a

The syntheses of the (4,1-benzoxazepin-3-ylidene)acetic acid derivatives **3** and **4**, which have an *n*-propyl and isobutyl group at the 1-position, are outlined in Chart 4. Acylation of racemic aminobenzylalcohol derivatives **5b**^{1b)} and **5c**^{1b)} with the acid chloride obtained from DL-*trans*-epoxysuccinic acid monoethyl ester afforded the epoxy-amide **6b**, **c**. Intramolecular cyclization of **6b**, **c** gave a diastereomeric mixture of α hydroxyacetic acid derivatives **10a**, **b** which were mesylated to give **11a** and **11b**. Subsequent treatment with cesium propionate, followed by chromatographic separation afforded **12a**, **b** and **13a**, **b**, respectively. The configurations of **12a**, **b** and **13a**, **b** were determined by comparison of the chemical shifts of signals due to olefin protons in the ¹H-NMR spectra of **9**, **12** and **13**. Alkaline hydrolysis of **12a**, **b** and **13a**, **b** yielded the carboxylic acid derivatives **3a**, **b** and **4a**, **b**, re-



ca. 1:2

Reagents: (a) MsCl, Et₃N, AcOEt; (b) 1) CH₃CH₂COOCs, DMF, 2) chromatographic separation.



Reagents: (a) DL-*trans*-Epoxysuccinic acid monoethyl ester, dichloromethylenedimethyliminium chloride, NaHCO₃, CH₂Cl₂; (b) K₂CO₃, EtOH; (c) MsCl, Et₃N, AcOEt; (d) 1) CH₃CH₂COOCs, DMF, 2) chromatographic separation; (e) 1 N NaOH, EtOH.

spectively.

Biological Results and Discussion

The compounds synthesized were evaluated for inhibition

Table 1. Squalene Synthase Inhibitory Activities of (\pm) -[7-Chloro-5-(2-
chlorophenyl)-2-oxo-1,5-dihydro-4,1-benzoxazepin-3-ylidene]aceticAcid
Derivatives (2—4) and (\pm) -(3,5-*trans*)-7-Chloro-5-(2-chlorophenyl)-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-acetic Acid Derivatives (1)



Compd.	Config.	R	IC ₅₀ (µм) ^{<i>a</i>)}	
			Rat enzyme	Human enzyme
2a	Ε	Pr ⁱ	0.31	0.17
2b	Ζ	Pr ⁱ	$>10 \ (21.6)^{b)}$	c)
3a	E	Pr ⁿ	0.076	0.098
3b	Ζ	Pr ⁿ	8.4	c)
4a	E	Bu^i	0.11	0.041
4b	Z	Bu^i	$>10 (14.5)^{b}$	c)
1a ^{d)}		Pr ⁱ	0.71	0.54
1b ^d)		Pr ⁿ	0.14	c)
1c ^{<i>d</i>})		Bu^{i}	0.061	0.034

a) IC_{50} values were determined by a single experiment run in duplicate. b) % Inhibition at 10^{-5} M. c) Not tested. d) Reference 1b.

of squalene synthase prepared from rat liver and human hepatoma (HepG2) cells. Inhibitory activities were measured according to the method of Cohen *et al.* with slight modification.⁹⁾

Inhibitory activities of the (4,1-benzoxazepin-3-ylidene)acetic acid derivatives 2—4 for squalene synthase are shown in Table 1. It can be seen that the inhibitory activity of the (E)-isomers 2a, 3a and 4a are of similar potency as the corresponding acetic acid derivatives 1a—c. On the other hand, the (Z)-isomers 2b, 3b and 4b were found to be only weakly active against rat enzyme.

In the case of the (*E*)-isomers, we assume that the double bonds holds the carboxyl groups in a position favorable for interaction with the enzyme, and conversely that the spatial positions of the carboxyl groups of the (*Z*)-isomers are unfavorable. Since the activities of **1a**—**c** and the (*E*)-isomers **2a**, **3a** and **4a** are almost identical, the active conformation of the 4,1-benzoxazepine-3-acetic acid derivatives is presumed be similar to the folded conformation of the (*E*)-(4,1-benzoxazepin-3-ylidene)acetic acid derivatives.¹⁰⁾ However, since all compounds described in this paper are racemates, it is not clear which enantiomer is the active inhibitor, and optical resolution of these compounds is currently in progress and will be reported in due course.

Conclusion

The (*E*)- and (*Z*)-(4,1-benzoxazepin-3-ylidene)acetic acid derivatives **2**—**4** were prepared and evaluated for their inhibition of squalene synthase. Interestingly, the (*E*)-isomers were much more potent than the (*Z*)-isomers against rat enzyme, and exhibited equivalent potency to the 4,1-benzoxazepine-3-acetic acid derivatives. Thus we assume that the active conformation of the 4,1-benzoxazepine-3-acetic acid derivatives is similar to the folded conformation of the (*E*)-(4,1-benzox-azepin-3-ylidene)acetic acid derivatives.

Chart 4

Experimental

All melting points were determined on a Yanagimoto micro melting point apparatus and are uncorrected. Proton nuclear magnetic resonance (¹H-NMR) spectra were recorded on a Varian GEMINI-200 (200 MHz) spectrometer (with tetramethylsilane as an internal standard). Infrared (IR) absorption spectra were recorded on a JASCO IR-810. [α]_D values were determined in the indicated solvents on a JASCO DIP-370 polarimeter. TLC analyses were carried out on Merck Kieselgel 60 F₂₅₄ plates. Elemental analyses were carried out by Takeda Analytical Laboratories, Ltd., and are within $\pm 0.4\%$ of the theoretical values. For column chromatography, Merck Kieselgel 60 (70—230 mesh ASTM) was used. Yields were not maximized. The following abbreviations are used: s=singlet, d=doublet, t=triplet, q= quartet, m=multiplet, br=broad.

Ethyl (2,3-*trans*)-3-[*N*-[4-Chloro-2-(2-chloro-α-hydroxybenzyl)phenyl]-*N*-isopropyl]carbamoyl]-2,3-epoxybutyrate (6a) A mixture of DL-*trans*-2,3-epoxysuccinic acid monoethyl ester³ (3.1 g, 19.3 mmol) and dichloromethylenedimethyliminium chloride (3.14 g, 19.3 mmol) in CH₂Cl₂ (90 ml) was stirred for 1 h with ice-cooling, followed by addition of (±)-5a (4.0 g, 12.9 mmol) and NaHCO₃ (5.42 g, 64.5 mmol). The solution was stirred for 1 h at 0 °C, washed with water, dried over MgSO₄, and then concentrated *in vacuo*. The residue was chromatographed [eluent: hexane–AcOEt (2 : 1)] to give 6a (5.6 g, 12.4 mmol, 96%) as a colorless powder. mp 143—146 °C (AcOEt–hexane). IR v_{max} (KBr) cm⁻¹: 3450, 3380 (NH, OH); 1750, 1660 (C=O). ¹H-NMR (CDCl₃) δ: 0.95—1.5 (9H, m), 2.5—3.9 (3H, m), 4.0—4.6 (3H, m), 6.25—6.6 (1H, m), 6.95—7.8 (7H, m). *Anal.* Calcd for C₂₂H₂₃Cl₂NO₅: C, 58.42; H, 5.13; N, 3.10. Found: C, 58.55; H, 5.31; N, 3.12.

Ethyl (3RS,5SR, aSR)-7-Chloro-5-(2-chlorophenyl)-1-isopropyl-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-glycolate (7a) and Ethyl (3RS, 5RS, aSR)-7-Chloro-5-(2-chlorophenyl)-1-isopropyl-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-glycolate (7b) A mixture of 6a (10.0 g, 22.1 mmol), K2CO3 (3.06 g, 2.21 mmol) and EtOH (200 ml) was stirred overnight at room temperature. The reaction mixture was diluted with water, extracted with AcOEt. The extract was washed with brine, dried over MgSO4, and then concentrated under reduced pressure. The residue was chromatographed [eluent: hexane-AcOEt (2:1)] to give a 1:1 mixture of 7a and 7b (7.2 g, 15.9 mmol, 72%). The mixture was recrystallized from AcOEthexane to give the $(3RS,5SR,\alpha SR)$ -isomer 7a (1.75 g, 3.87 mmol, 24%) as colorless prisms. mp 188—189 °C (AcOEt-hexane). ¹H-NMR (CDCl₃) δ : 1.29 (3H, t, J=7.1 Hz), 1.30 (3H, d, J=7.0 Hz), 1.56 (3H, d, J=7.0 Hz), 4.07 (1H, d, J=9.0 Hz), 4.19 (1H, d, J=4.8 Hz), 4.26 (2H, dq, J=7.1, 1.8 Hz), 4.5-4.65 (1H, m), 4.7-4.9 (1H, m), 6.04 (1H, s), 6.54 (1H, d, J=2.6 Hz), 7.2-7.8 (6H, m).

Mother liquid was concentrated *in vacuo*. The residue was recrystallized from AcOEt–hexane to give the $(3RS,5RS,\alpha SR)$ -isomer **7b** (1.2 g, 2.65 mmol, 17%) as colorless prisms. mp 141—142 °C (AcOEt–hexane). ¹H-NMR (CDCl₃) δ : 0.4—0.6 (3H, m), 1.24 (3H, d, *J*=7.4 Hz), 1.26 (3H, t, *J*=7.4 Hz), 3.95—4.25 (2H, m), 4.23 (2H, q, *J*=7.4 Hz), 4.38 (1H, d, *J*=5.0 Hz), 4.4—4.6 (1H, m), 6.09 (1H, s), 7.1—7.85 (7H, m).

Ethyl $(3RS, 5SR, \alpha SR) - \alpha$ -Methanesulfonyloxy-7-chloro-5-(2chlorophenyl)-1-isopropyl-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-acetate (8a) and Ethyl (3RS,5RS, aSR)-a-Methanesulfonyloxy-7chloro-5-(2-chlorophenyl)-1-isopropyl-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-acetate (8b) MsCl (0.13 ml, 1.72 mmol) and Et₃N (0.24 ml, 1.72 mmol) was added to a solution of 7a (0.6g, 1.33 mmol) in AcOEt (30 ml) with ice-cooling. After stirring for 1 h, the reaction mixture was washed with brine, dried over MgSO4, and then concentrated. The residue was chromatographed [eluent: hexane-CH₂Cl₂-AcOEt (10:5:1)] to give 8a (0.63 g, 1.19 mmol, 89%) as colorless prisms. mp 159-160 °C (hexane-AcOEt). IR v_{max} (KBr) cm⁻¹: 1745, 1680 (C=O). ¹H-NMR (CDCl₃) δ : 1.31 (3H, t, J=7.1 Hz), 1.31 (3H, d, J=7.0 Hz), 1.56 (3H, d, J=7.0 Hz), 3.17 (3H, s), 4.18 (1H, d, J=8.0 Hz), 4.32 (2H, q, J=7.1 Hz), 4.7-4.9 (1H, m), 5.37 (1H, d, J=8.0 Hz), 6.04 (1H, s), 6.50 (1H, d, J=2.4 Hz), 7.2-7.6 (6H, m). Anal. Calcd for C23H25Cl2NO7S: C, 52.08; H, 4.75; N, 2.64. Found: C, 52.25; H, 4.80; N, 2.74.

Compound **8b** (0.82 g, 1.55 mmol, 88%) was prepared from **7b** (0.80 g, 1.77 mmol) in a similar manner as described for the preparation of **8a** as a colorless prisms. mp 143—144 °C (AcOEt–hexane). IR v_{max} (KBr) cm⁻¹: 1750, 1670 (C=O). ¹H-NMR (CDCl₃) δ : 0.5—0.8 (3H, m), 1.26 (3H, d, *J*= 6.8 Hz), 1.29 (3H, t, *J*=7.0 Hz), 3.16 (3H, s), 4.28 (2H, q, *J*=7.0 Hz), 4.1—4.35 (1H, m), 4.45—4.6 (1H, m), 5.1—5.4 (1H, m), 6.09 (1H, s), 7.1—7.75 (7H, m). *Anal.* Calcd for C₂₃H₂₅Cl₂NO₇S: C, 52.08; H, 4.75; N, 2.64. Found: C, 52.38; H, 4.89; N, 2.77.

Ethyl (E)-[7-Chloro-5-(2-chlorophenyl)-1-isopropyl-2-oxo-1,5-dihydro-4,1-benzoxazepin-3(2H)-ylidene]acetate (9a) and Ethyl (Z)-[7-Chloro-5(2-chlorophenyl)-1-isopropyl-2-oxo-1,5-dihydro-4,1-benzoxazepin-3(2*H*)ylidene]acetate (9b) A mixture of propionic acid (0.23 g, 3.11 mmol) and Cs_2CO_3 (0.34 g, 1.04 mmol) in MeOH (10 ml) was stirred for 30 min at room temperature, and then concentrated under reduced pressure to give cesium propionate as a colorless powder. A mixture of **8a** (1.0 g, 1.89 mmol) and cesium propionate in *N*,*N*-dimethylformamide (DMF, 10 ml) was stirred for 2 h at 80 °C, diluted with water, and extracted with AcOEt. The extract was washed with 1 \times HCl and saturated NaHCO₃, dried over MgSO₄, and then concentrated *in vacuo*. The residue was chromatographed [eluent: hexane–AcOEt (10 : 1 then 5 : 1)] to give **9b** (0.31 g, 0.714 mmol, 38%) as colorless prisms from the first fraction and **9a** (0.13 g, 0.299 mmol, 16%) as a colorless prisms from the second fraction.

9a: mp 143—144 °C (Et₂O–hexane). IR v_{max} (KBr) cm⁻¹: 1710, 1670 (C=O), 1650 (C=C). ¹H-NMR (CDCl₃) δ : 1.23 (3H, t, *J*=7.2 Hz), 1.34 (3H, d, *J*=7.0 Hz), 1.66 (3H, d, *J*=7.0 Hz), 4.05—4.3 (2H, m), 4.9—5.1 (1H, m), 5.41 (1H, s), 6.49 (1H, s), 6.53 (1H, s), 7.3—7.6 (5H, m), 7.78 (1H, d, *J*=6.8 Hz). *Anal.* Calcd for C₂₂H₂₁Cl₂NO₄: C, 60.84; H, 4.87; N, 3.22. Found: C, 60.59; H, 4.84; N, 3.30.

9b: mp 194—195 °C (Et₂O–hexane). IR v_{max} (KBr) cm⁻¹: 1700, 1650 (C=O), 1630 (C=C). ¹H-NMR (CDCl₃) δ : 1.27 (3H, t, *J*=7.0 Hz), 1.30 (3H, d, *J*=6.8 Hz), 1.61 (3H, d, *J*=6.8 Hz), 4.05—4.3 (2H, m), 4.7—4.95 (1H, m), 5.48 (1H, s), 6.52 (1H, s), 6.57 (1H, d, *J*=2.2 Hz), 7.3—7.6 (5H, m), 8.16 (1H, d, *J*=7.4 Hz). *Anal*. Calcd for C₂₂H₂₁Cl₂NO₄: C, 60.84; H, 4.87; N, 3.22. Found: C, 60.76; H, 4.99; N, 3.32.

8b (1.0 g, 1.89 mmol) was converted to a mixture of **9a** and **9b** in a similar manner as described above. The mixture was chromatographed [eluent: hexane–AcOEt (10:1-5:1)] to give **9b** (0.33 g, 0.760 mmol, 40%) as colorless prisms from the first fraction and **9a** (0.17 g, 0.391 mmol, 21%) as a colorless prisms from the second fraction.

(E)-[7-Chloro-5-(2-chlorophenyl)-1-isopropyl-2-oxo-1,5-dihydro-4,1benzoxazepin-3(2H)-ylidene]acetic Acid (2a) and (Z)-[7-Chloro-5-(2chlorophenyl)-1-isopropyl-2-oxo-1,5-dihydro-4,1-benzoxazepin-3(2H)ylidene]acetic Acid (2b) One normal NaOH (5 ml) was added to a solution of 9a (0.16 g, 0.368 mmol) in MeOH (20 ml). The mixture was stirred for 1 h at room temperature, diluted with water, acidified, and extracted with AcOEt. The extract was washed with brine, dried over MgSO₄, and then concentrated under reduced pressure to give 2a (0.14 g, 0.345 mmol, 94%) as colorless plates.

Compound **2b** (0.26 g, 0.640 mmol, 93%) was prepared from **9b** (0.3 g, 0.691 mmol) in a similar manner as described for the preparation of **2a** as a colorless plates.

2a: mp 209—214 °C (dec.) (AcOEt–hexane). IR v_{max} (KBr) cm⁻¹: 1720, 1650 (C=O), 1630 (C=C). ¹H-NMR (CDCl₃) δ : 1.34 (3H, d, J=7.0 Hz), 1.62 (3H, d, J=7.0 Hz), 4.8—5.0 (1H, m), 5.41 (1H, s), 6.51 (1H, s), 6.54 (1H, d, J=1.8 Hz), 7.3—7.6 (5H, m), 7.78 (1H, d, J=7.0 Hz). Anal. Calcd for C₂₀H₁₇Cl₂NO₄: C, 59.13; H, 4.22; N, 3.45. Found: C, 58.97; H, 4.24; N, 3.42.

2b: mp 234—239 °C (dec.) (AcOEt–hexane). IR v_{max} (KBr) cm⁻¹: 1670 (C=O), 1655 (C=C). ¹H-NMR (CDCl₃) δ : 1.31 (3H, d, J=7.0 Hz), 1.61 (3H, d, J=7.0 Hz), 4.7—4.9 (1H, m), 5.48 (1H, s), 6.54 (1H, s), 6.57 (1H, d, J=2.2 Hz), 7.3—7.6 (5H, m), 8.05 (1H, d, J=7.8 Hz). *Anal.* Calcd for C₂₀H₁₇Cl₂NO₄: C, 59.13; H, 4.22; N, 3.45. Found: C, 59.02; H, 4.21; N, 3.37.

Ethyl (2,3-*trans*)-3-[*N*-[4-Chloro-2-(2-chloro- α -hydroxybenzyl)phenyl]-*N*-propyl]carbamoyl]-2,3-epoxybutyrate (6b) and Ethyl (2,3-*trans*)-3-[*N*-[4-Chloro-2-(2-chloro- α -hydroxybenzyl)phenyl]-*N*-isobutyl]carbamoyl]-2,3-epoxybutyrate (6c) Compound 6b (5.3 g, 11.7 mmol, 91%) and 6c (14.0 g, 30.0 mmol, 97%) were prepared from (±)-5b (4.0 g, 12.9 mmol) and (±)-5c (10.0 g, 30.8 mmol) in a similar manner as described for the preparation of 6a.

6b: An oil. IR v_{max} (neat) cm⁻¹: 1745, 1660 (C=O). ¹H-NMR (CDCl₃) δ : 0.75—1.05 (3H, m), 1.1—1.8 (5H, m), 2.6—3.9 (4H, m), 4.0—4.5 (3H, m), 6.1—6.4 (1H, m), 6.9—7.9 (7H, m).

6c: An oil. IR v_{max} (neat) cm⁻¹: 3390 (OH); 1745, 1660 (C=O). ¹H-NMR (CDCl₃) δ : 0.75—1.05 (6H, m), 1.15—1.4 (3H, m), 1.7—2.0 (1H, m), 2.7—3.3 (2H, m), 3.3—3.9 (2H, m), 4.0—4.45 (3H, m), 6.1—6.4 (1H, m), 7.0—7.8 (7H, m).

Ethyl (3,5-*trans*)-7-Chloro-5-(2-chlorophenyl)-2-oxo-1-propyl-1,2,3,5tetrahydro-4,1-benzoxazepine-3-glycolate (10a) and Ethyl (3,5-*trans*)-7-Chloro-5-(2-chlorophenyl)-1-isobutyl-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-glycolate (10b) A mixture of 6c (13.0 g, 27.9 mmol) and K_2CO_3 (3.85 g, 27.9 mmol) in EtOH (150 ml) was stirred overnight at room temperature, diluted with water, and extracted with AcOEt. The extract was washed with brine, dried over MgSO₄, and then concentrated. The residue was chromatographed [eluent: hexane–AcOEt (2:1)] to give **10b** (a mixture of diastereomers) (7.0 g, 15.0 mmol, 54%) as colorless crystals.

10b: IR v_{max} (KBr) cm⁻¹: 3400 (OH); 1740, 1670, 1640 (C=O). ¹H-NMR (CDCl₃) δ : 0.7—1.1 (6H, m), 1.2—1.4 (5H, m), 1.7—2.1 (1H, m), 3.35—4.0 (2H, m), 4.0—4.8 (5H, m), 6.15, 6.19 (1H, each s, *ca.* 1 : 1), 6.55, 6.96 (1H, each d, J=2.4 Hz), 7.15—7.8 (6H, m). *Anal.* Calcd for C₂₃H₂₅Cl₂NO₅: C, 59.24; H, 5.40; N, 3.00. Found:C, 59.14; H, 5.37; N, 2.98.

Compound 10a (2.3 g, 5.08 mmol, 43%) was prepared from **6b** (5.3 g, 11.7 mmol) in a similar manner as described for the preparation of **10b**.

10a: An oil. IR v_{max} (KBr) cm⁻¹: 3380 (OH); 1745, 1650 (C=O). ¹H-NMR (CDCl₃) δ : 0.7—1.05 (3H, m), 1.15—1.4 (3H, m), 1.45—1.8 (2H, m), 3.4—3.9 (2H, m), 4.0—4.7 (4H, m), 6.08, 6.10 (1H, each s), 6.5—7.8 (7H, m). *Anal.* Calcd for C₂₂H₂₃Cl₂NO₅: C, 58.42; H, 5.12; N, 3.10. Found : C, 58.40; H, 5.23; N, 3.01.

Ethyl α -Methanesulfonyloxy-(3,5-*trans*)-7-chloro-5-(2-chlorophenyl)-2-oxo-1-propyl-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-acetate (11a) and Ethyl α -Methanesulfonyloxy-(3,5-*trans*)-7-chloro-5-(2-chlorophenyl)-1isobutyl-2-oxo-1,2,3,5-tetrahydro-4,1-benzoxazepine-3-acetatel (11b) Compounds 11a (1.9 g, 3.58 mmol, 81%) and 11b (2.0 g, 3.67 mmol, 95%) were prepared from 10a (2.0 g, 4.42 mmol) and 10b (1.8 g, 3.86 mmol) in a similar manner as described for the preparation of 8a.

11a: An oil. IR v_{max} (neat) cm⁻¹: 1750, 1670 (C=O). ¹H-NMR (CDCl₃) δ : 0.7—1.1 (3H, m), 1.2—1.4 (3H, m), 1.5—1.85 (2H, m), 3.07, 3.16 (3H, each s), 3.4—3.8 (2H, m), 4.2—5.45 (4H, m), 6.07, 6.10 (1H, each s), 6.5—7.7 (7H, m).

11b: A mixture of crystals and an oil. IR v_{max} (Nujol) cm⁻¹: 1765, 1670, 1630 (C=O). ¹H-NMR (CDCl₃) δ : 0.75—1.1 (6H, m), 1.25—1.4 (3H, m), 1.7—2.1 (1H, m), 2.98, 3.16 (3H, each s), 3.35—3.6 (1H, m), 3.9—5.5 (5H, m), 6.18 (1H, s), 6.5—7.85 (7H, m). *Anal.* Calcd for $C_{24}H_{27}Cl_2NO_7S$: C, 52.95; H, 5.00; N, 2.57. Found: C, 52.92; H, 5.07; N, 2.72.

Ethyl (*E*)- and (*Z*)-[7-Chloro-5-(2-chlorophenyl)-2-oxo-1-propyl-1,5dihydro-4,1-benzoxazepin-3(2*H*)-ylidene]acetate (12a, b) and Ethyl (*E*)and (*Z*)-[7-Chloro-5-(2-chlorophenyl)-1-isobutyl-2-oxo-1,5-dihydro-4,1benzoxazepine-3(2*H*)-ylidene]acetate (13a, b) Compounds 12a, b and 13a, b were prepared from 11a (1.9 g, 3.58 mmol) and 11b (2.0 g, 3.67 mmol) in a similar manner as described for the preparation of 9a, b.

12a (Polar): Prisms. Yield 0.20 g (0.461 mmol, 13%). mp 114—115 °C (Et₂O–hexane). IR v_{max} (KBr) cm⁻¹: 1715, 1670 (C=O), 1640 (C=C). ¹H-NMR (CDCl₃) δ : 1.00 (3H, t, *J*=7.4 Hz), 1.27 (3H, t, *J*=7.1 Hz), 1.5—1.9 (2H, m), 3.5—3.75 (1H, m), 4.13 (2H, q, *J*=7.1 Hz), 4.3—4.55 (1H, m), 5.45 (1H, s), 6.49 (1H, s), 6.53 (1H, d, *J*=1.8 Hz), 7.3—7.6 (5H, m), 7.78 (1H, d, *J*=6.6 Hz). *Anal.* Calcd for C₂₂H₂₁Cl₂NO₄: C, 60.84; H, 4.87; N, 3.22. Found: C, 60.85; H, 5.17; N, 3.13.

12b (Less Polar): Prisms. Yield 0.61 g (1.40 mmol, 39%). mp 171— 172 °C (Et₂O–hexane). IR v_{max} (KBr) cm⁻¹: 1710, 1660 (C=O), 1630 (C=C). ¹H-NMR (CDCl₃) δ : 0.98 (3H, t, J=7.4 Hz), 1.28 (3H, t, J=7.1 Hz), 1.6—1.85 (2H, m), 3.45—3.7 (1H, m), 4.1—4.3 (2H, m), 4.4—4.6 (1H, m), 5.50 (1H, s), 6.53 (1H, s), 6.57 (1H, d, J=2.2 Hz), 7.2—7.6 (5H, m), 8.17 (1H, d, J=7.2 Hz). *Anal*. Calcd for C₂₂H₂₁Cl₂NO₄: C, 60.84; H, 4.87; N, 3.22. Found: C, 60.65; H, 4.73; N, 3.15.

13a (Polar): An oil. Yield 0.29 g (0.647 mmol, 18%). IR v_{max} (Neat) cm⁻¹: 1710, 1670 (C=O), 1645 (C=C). ¹H-NMR (CDCl₃) δ : 0.95 (3H, d, J=6.6 Hz), 1.07 (3H, d, J=6.6 Hz), 1.9—2.2 (1H, m), 3.57 (1H, dd, J=14.0, 5.8 Hz), 4.0—4.25 (2H, m), 4.37 (1H, dd, J=14.0, 8.6 Hz), 5.44 (1H, s), 6.54 (1H, d, J=1.8 Hz), 6.57 (1H, s), 7.3—7.9 (6H, m).

13b (Less Polar): Prisms. Yield 0.89 g (1.99 mmol, 54%). mp 181— 182 °C (Et₂O-hexane). IR v_{max} (KBr) cm⁻¹: 1715 (C=O), 1650 (C=C). ¹H-NMR (CDCl₃) δ : 0.94 (3H, d, J=6.6 Hz), 1.01 (3H, d, J=6.6 Hz), 1.85—2.1 (1H, m), 3.43 (1H, dd, J=13.8, 5.6 Hz), 4.05—4.25 (2H, m), 4.45 (1H, dd, J=13.8, 8.6 Hz), 5.51 (1H, s), 6.57 (1H, d, J=2.4 Hz), 6.61 (1H, s), 7.2—7.6 (5H, m), 8.19 (1H, d, J=8.0 Hz). Anal. Calcd for C₂₃H₂₃Cl₂NO₄: C, 61.62; H, 5.17; N, 3.12. Found: C, 61.54; H, 5.30; N, 3.09.

(*E*)- and (*Z*)-[7-Chloro-5-(2-chlorophenyl)-2-oxo-1-propyl-1,5-dihydro-4,1-benzoxazepin-3-ylidene]acetic Acid (3a,b; Table 1) and Ethyl (*E*)- and (*Z*)-[7-Chloro-5-(2-chlorophenyl)-1-isobutyl-2-oxo-1,5-dihydro-4,1-benzoxazepin-3-ylidene]acetic Acid (4a,b; Table 1) Compounds 3a, b and 4a, b were prepared from 12a (0.17 g, 0.391 mmol), 12b (0.30 g, 0.691 mmol), 13a (0.28 g, 0.625 mmol) and 13b (0.30 g, 0.669 mmol) in a similar manner as described for the preparation of 2a, b.

3a: Prisms. Yield 0.15 g (0.369 mmol, 94%). mp 192—194 °C (dec.) (AcOEt–hexane). IR v_{max} (KBr) cm⁻¹: 1715, 1645 (C=O), 1635 (C=C). ¹H-NMR (CDCl₃) δ : 1.01 (3H, t, J=7.4 Hz), 1.6—1.9 (2H, m), 3.6—3.8 (1H, m), 4.25—4.5 (1H, m), 5.47 (1H, s), 6.51 (1H, s), 6.54 (1H, d, J=2.2 Hz), 7.3—7.6 (5H, m), 7.80 (1H, d, J=6.6 Hz). Anal. Calcd for

 $C_{20}H_{17}Cl_2NO_4\!\!:$ C, 59.13; H, 4.22; N, 3.45. Found: C, 58.88; H, 4.34; N, 3.27.

3b: Prisms. Yield 0.23 g (0.566 mmol, 82%). mp 208—210 °C (dec.) (AcOEt-hexane). IR v_{max} (KBr) cm⁻¹: 1690, 1655 (C=O), 1635 (C=C). ¹H-NMR (CDCl₃) δ : 0.98 (3H, t, J=7.4 Hz), 1.6—1.9 (2H, m), 3.4—3.7 (1H, m), 4.4—4.6 (1H, m), 5.51 (1H, s), 6.56 (1H, s), 6.58 (1H, d, J=2.4 Hz), 7.2—7.6 (5H, m), 8.07 (1H, d, J=7.0 Hz). Anal. Calcd for C₂₀H₁₇Cl₂NO₄: C, 59.13; H, 4.22; N, 3.45. Found: C, 59.20; H, 4.39; N, 3.32.

4a: Prisms. Yield 0.25 g (0.595 mmol, 95%). mp 207—209 °C (dec.) (AcOEt–hexane). IR $v_{\rm max}$ (KBr) cm⁻¹: 1720 (C=O), 1645 (C=C). ¹H-NMR (CDCl₃) δ : 0.97 (3H, d, *J*=6.7 Hz), 1.08 (3H, d, *J*=6.6 Hz), 1.95—2.2 (1H, m), 3.63 (1H, dd, *J*=13.9, 5.6 Hz), 4.23 (1H, dd, *J*=13.9, 8.8 Hz), 5.46 (1H, s), 6.55 (1H, d, *J*=2.2 Hz), 6.58 (1H, s), 7.3—7.9 (6H, m). *Anal.* Calcd for C₂₁H₁₉Cl₂NO₄: C, 60.01; H, 4.56; N, 3.33. Found: C, 59.95; H, 4.64; N, 3.45.

4b: Prisms. Yield 0.25 g (0.595 mmol, 89%). mp 236—238 °C (dec.) (AcOEt–hexane). IR $v_{\rm max}$ (KBr) cm⁻¹: 1690, 1655 (C=O), 1630 (C=C). ¹H-NMR (CDCl₃) δ : 0.95 (3H, d, *J*=6.6 Hz), 1.02 (3H, d, *J*=6.6 Hz), 1.85—2.1 (1H, m), 3.45 (1H, dd, *J*=13.8, 5.8 Hz), 4.46 (1H, dd, *J*=13.8, 8.6 Hz), 5.52 (1H, s), 6.57 (1H, d, *J*=2.2 Hz), 6.64 (1H, s), 7.2—7.6 (5H, m). *Anal.* Calcd for C₂₁H₁₉Cl₂NO₄: C, 60.01; H, 4.56; N, 3.33. Found: C, 60.16; H, 4.56; N, 3.50.

Animals and Materials Animals were supplied by Clea, Japan, Inc. unless otherwise mentioned. *RS*-[2-¹⁴C]Mevalonolactone and [1-³H]farnesyl pyrophosphate were purchased from New England Nuclear. [2-¹⁴C]mevalonic acid was synthesized from [2-¹⁴C]Mevalonolactone by saponification with potassium hydroxide. [2-¹⁴C]Sodium acetate was purchased from Amersham. Farnesyl pyrophosphate was synthesized by the method described by V. J. Davisson and coworkers¹¹⁾ (Nemoto & Co.). HepG2 cells were supplied by The American Type Culture Collection (ATCC). Fetal bovine serum (FBS) and Dulbecco's modified Eagle's medium (DMEM) were purchased from GIBCO. Human lipoprotein deficient serum (human LPDS) was purchased from Sigma. All other reagents were supplied by Wako Pure Chemical Industries.

Preparation of Rat Squalene Synthase An Sprague-Dawley (SD) male rat (6-week old) was killed by bleeding, and its liver was excised. About 10 g of the liver was washed with a saline solution cooled with ice, which was then homogenized in 15 ml of an ice-cooled buffer solution [100 mM potassium phosphate (pH 7.4), 15 mM nicotinamide, 2 mM MgCl₂], followed by centrifugation for 20 min at $10000 \times g$ (4 °C). The supernatant layer was separated and subjected to further centrifugation for 90 min at $105000 \times g$ (4 °C). The sediment was then suspended in an ice-cooled 100 mM potassium phosphate buffer solution (pH 7.4), which was again subjected to centrifugation for 90 min at $105000 \times g$ (4 °C). The sediment thus obtained (microsome fraction) was suspended in an ice-cooled 100 mM potassium phosphate buffer (pH 7.4) (about 40 mg/ml protein concentration, determined using Bicinchoninic acid (BCA) protein assay kit of Pierce Co., Ltd.). This suspension was used as the enzyme solution.

Preparation of Human Squalene Synthase HepG2 cells (about 1×10^9 cells) obtained by incubation (37 °C in the presence of 5% CO₂) in a DMEM contains 10% FBS, penicillin G (100 units/ml) and streptomycin ($10 \,\mu g/ml$) were suspended in 10 ml of ice-cooled buffer solution [100 mM potassium phosphate buffer (pH 7.4), 30 mM nicotinamide and 2.5 mM MgCl₂]. The cells were crashed by means of ultrasonication (for 30 s, twice). From the sonicate thus obtained, the microsome fraction was obtained by the same procedure as in preparation of rat-derived enzyme, which was suspended in an ice-cooled 100 mM potassium phosphate buffer (pH 7.4) (about 4 mg/ml protein concentration). This suspension was used as the enzyme solution.

Assay of Squalene Synthase Inhibitory Activity Squalene synthase activity was monitored by the formation of [³H]squalene from [1-³H]farnesyl pyrophosphate. Fifty microliters of assay mixture included $5 \mu M$ [1-³H]farnesyl pyrophosphate (25 μ Ci/mol), 1 mM NADPH, 5 mM MgCl₂, 6 mM glutathione, 100 mM buffer solution of potassium phosphate (pH 7.4), the test compound dissolved in dimethyl sulfoxide (DMSO) (a final concentration of DMSO was 2%) and enzyme solution prepared from rat or HepG2 cells (protein content 0.8 μ g). The assay ran 45 min at 37 °C and stopped by adding 150 μ l of CHCl₃–MeOH (1 : 2) containing 0.2% cold squalene as carrier. Aqueous solution of 3 N NaOH (50 μ M) and CHCl₃ (50 μ M) were added to the mixture. The chloroform layer containing the reaction mixture having squalene as the principal component and 3 ml of toluene-based liquid scintillator were mixed, and its radioactivity was determined by means of a liquid scintillation counter. The squalene synthase inhibitory activity was expressed in terms of the concentration of the test compound inhibiting by 50% the radioactivity taken into the chloroform layer (IC₅₀, molar concentration (M)).

Single-Crystal X-Ray Analysis. 7a: Unit cell parameters [a=11.245 (1) Å, b=11.746 (1) Å, c=9.067 (1) Å, $\alpha=107.26$ (1)°, $\beta=98.64$ (1)°, $\gamma=74.94$ (1)°], Z=2, space group P1, *R*-factor=0.080.

7b: Unit cell parameters [a=10.854 (1) Å, b=11.083 (1) Å, c=9.807 (1) Å, $\alpha=108.20$ (1)°, $\beta=96.53$ (1)°, $\gamma=94.13$ (1)°], Z=2, space group P1, *R*-factor=0.083.

2a: Unit cell parameters [a=10.018 (1) Å, b=11.304 (1) Å, c=8.508 (1) Å, α =93.87 (1)°, β =98.23 (1)°, γ =91.83 (1)°], Z=2, space group *P*1, *R*-factor=0.063.

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References and Notes

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